

Determination of Cytotoxicity of Zinc 2-Bromobenzoate with Nicotinamide and N,N'-Diethylnicotinamide Complexes

Giray Buğra AKBABA

Kafkas University, Faculty of Engineering-Architecture, Department of Bioengineering, 36100, Kars, Turkey.

Araştırma Makalesi
Research Article

Kimya
Chemistry

Geliş Tarihi/Received
23.11.2020

Kabul Tarihi/Accepted
30.12.2020

Abstract: Benzoic acid and its derivatives and their metal complexes, which have antimicrobial, anticancer, antituberculosis and antioxidant properties, are biologically active molecules. Although there are many studies on the biological activity of these compounds, studies on the determination of their toxicity are limited. In the presented study, the cytotoxic properties of the previously synthesized diaquabis(2-bromobenzoato- κ O)bis(nicotinamide- κ N1)zinc(II) (ZnBrBANA) and diaquabis(2-bromobenzoato- κ O)bis(N,N'-diethylnicotinamide- κ N1)zinc(II) (ZnBrBADENA) complexes were investigated. The 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay, a colorimetric method, was used to determine the cytotoxicity of complexes on human peripheral lymphocyte cells. Besides, cytotoxicity of dimethylsulfoxide (DMSO) which is crystal solvent and 2-bromobenzoic acid (BrBA), nicotinamide (NA), and N,N'-diethylnicotinamide (DNA) which are starting compounds of the complexes was also evaluated. According to the results of MTT method, It has been determined that both complexes and starting components except BrBA cause cytotoxicity on lymphocyte cells at the concentration range of 62.5-500 ppm. In addition, it was determined that the BrBA and DMSO at the same concentration range do not show any cytotoxic effect on lymphocyte cells. It was observed that the synthesized complexes were more toxic at each concentration than the starting components. Therefore, the toxic effects of the complexes used as drug active ingredients should be followed up with new studies.

Keywords: Cytotoxicity, MTT assay, 2-Bromobenzoic acid, Zinc Complex

1. INTRODUCTION

In recent years, the cytotoxic and genotoxic properties of metal complexes and their roles in biological processes have been investigated (Mjos and Orvig, 2014). One of the most important reasons for this is the ability of metal drugs to target to DNA. The biological activity of these compounds depends on their structure and the ligands included. But crystal engineering must take into account any interaction, both weak and strong, to clarify molecular architecture and crystal packaging (Lu et al., 2019). Investigation of the relationship between the structure of compounds and their biological activities makes it possible to synthesize new drugs suitable for the purpose (Balan et al., 2020). Zinc complexes attract attention with their crystal structures as well as their biological and physical properties. In recent years, many zinc complexes containing biologically active ligands have been synthesized by different research groups and

their crystal structures have been investigated (Hökelek et al., 2009; Özbek et al., 2019; Pucci et al., 2013; Sertçelik et al. 2012; 2018; Taşdemir et al., 2016). Zinc (II) complexes are used as DNA binders, tumor photosensitizers, antidiabetic, antifungal, antioxidant and antibacterial agents (Liguori et al., 2010; Pucci et al., 2013). Moreover, in recent studies, Zn (II) complexes have reported that they can be anticancer agents due to their low toxicity and less side effects (Liguori et al., 2010; Pucci et al., 2013).

Metal complexes of benzoic acid and its derivatives have great abilities in various fields (Bakhtiar and Ochiai, 1999; Heine and Müller-Buschbaum, 2013; Krishna, 2015; Li et al., 2013; Rimoldi et al., 2017; Zhu et al., 2017). For the rational design, construction of their supramolecular architecture and properties, auxiliary ligands used as well as the main ligands are also important (Jozef et al., 2016). Used as auxiliary ligands, N-donor heterocyclic compounds are a component of various vitamins and drugs and play an important role in many biological systems. Nicotinamide, the body form of vitamin B3, is a N-donor ligand used in the treatment of some skin diseases. Another N-donor ligand with a similar structure is N,N'-diethylnicotinamide, which is used as a respiratory stimulator in medicine (Cavicchi, 1959; Krajníková et al., 2011). While the crystal structure of metal arylcarboxylates with different co-ligands changes, it is found that the biological activities and toxicity profiles of the complexes change with the change of these ligands (Wang et al., 2020). Drug active ingredients are carried by the blood tissue in the body. Therefore, these active substances first come into contact with blood tissue cells. Therefore, the cytotoxic effects of the two complexes used in this study on lymphocyte cells were investigated. The aim of this study, to determine the cytotoxicity effects of previously synthesized zinc 2-bromobenzoate complexes with nicotinamide and N,N'-diethylnicotinamide were used by MTT test method on lymphocyte cells.

2. MATERIAL AND METHOD

2.1. Materials

The two complexes used in the study were a previously synthesized and their structures were determined (Hökelek et al. 2009a; 2009b). The structures of the complexes were given in Figure 1 and 2. Phosphate Buffered Saline, Antibiotic Antimycotic Solution, L-Glutamine solution, Histopaque-1077 and Dimethylsulfoxide (Sigma-Aldrich), BIOAMF-1 medium and BIOAMF-1 supplement (Biological Industries) and MTT Cell Proliferation Assay Kit (Cayman Chemical) were purchased commercially. Solutions of the complexes at concentrations of 62.5, 125, 250 and 500 ppm were prepared in DMSO. Nüve Steamart OT 40L autoclave, Nüve BM

101 Water bath, ISOLAB vortex mixer, HETTICH EBA 200 centrifuge device, Panasonic MCO-170AICUVH-PE CO₂ Incubator and BioTek Epoch Spectrophotometer were used.

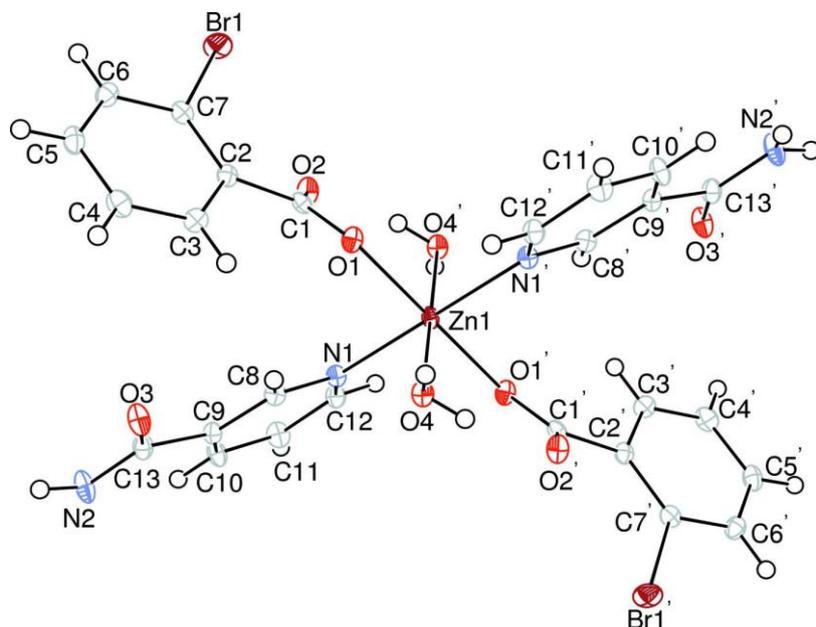


Figure 1. The structure of diaquabis(2-bromobenzoato- κ O)bis(nicotinamide- κ N¹)zinc(II)(Hökelek et al., 2009a)

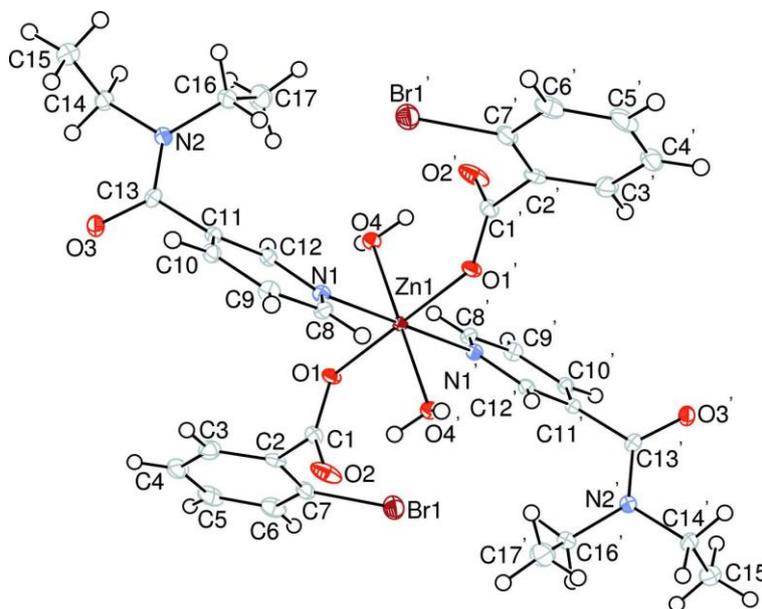


Figure 2. The structure of diaquabis(2-bromobenzoato- κ O)bis(N,N'-diethylnicotinamide- κ N¹)zinc(II)(Hökelek et al., 2009b)

2.2. Method

Lymphocyte cells to be used in experiments were isolated from a human whole blood sample (Öztürk, 2019) and cell count was done using Thoma slide. The culture medium was

prepared from the a mixture of amnion cell culture medium (75 mL), supplement (15 mL), penicillin + streptomycin + amphotericin B (Antibiotic Antimycotic Solution) (1.5 mL) and L-glutamine (2 mL) into a sterile tube. After that culture medium (100 μ L) and cell suspension (100 μ L) (50000 cells/well) were added to the 96-well plates, respectively. The cells were incubated to proliferate and adhere to the surface for 24 hours in 5 % CO₂ incubator at 37 °C. The solutions of the complexes at 500, 250, 125 and 62.5 ppm concentrations were prepared in DMSO. After 24 h incubation, 100 μ L aliquots of different concentrations of the complexes were added to the wells. The cell cultures were incubated at 37 °C for a day in the incubator. 24 hours later 10 μ L of MTT reagent was added to each well and the plates were gently mixed on the shaker. Incubation was continued for 3 hours. Formed formazan crystals were seen in the bottom of the wells. Then, the medium in the well was completely removed and 200 μ L DMSO was added to each well to dissolve the formazan crystals. It was kept in the incubator at 37 °C for 24 hours. At the end of the incubation, the absorbance values were recorded by UV-Vis spectrophotometer at 570 nm.

2.3. Statistical analysis

The data obtained from tests were analyzed with IBM SPSS statistics for Windows package program (v.18.0, IBM Corp., Armonk, New York, USA). Two-way ANOVA (Tukey) was used to evaluate whether any treatment significantly differed from the control or each other's. Statistically significance level was accepted at % 95 (p<0.05).

3. RESULTS

3.1. MTT Assay

MTT test is one of the most important pre-screening methods to investigate cell proliferation and anticancer activity of natural products and synthetic materials in the search for new drugs (Jamalian et al., 2011; Mosmann, 1983; Teixeira et al., 2007). In this context, MTT test's results of the complexes, solvent and starting components were assessed. The values of absorbance were recorded at 570 nm by spectrophotometer. The change in cell viability was compared with the cell control group and percentage of inhibition values were calculated according to the following equation (1). The values were given in Table 1 and Figure 3.

$$\text{Inhibition percentage (\%)} = \frac{CV}{CVCC} \times 100 \quad (1)$$

(CV= Cell viability at the test concentrations and CVCC = Cell viability in cell control)

Table 1. Percentage of Cell viability (CV) and Cell death (CD) values at the test concentrations

		500 ppm	250 ppm	125 ppm	62,5 ppm
Cell Control	CV	100.00	100.00	100.00	100.00
	CD	–	–	–	–
DMSO	CV	102.24	102.24	102.24	102.24
	CD	–	–	–	–
ZnBrBANA	CV	56.50	63.21	63.19	63.30
	CD	43.50	36.79	36.81	36.70
ZnBrBADENA	CV	66.03	76.23	73.32	86.01
	CD	33.97	23.77	26.68	13.99
NA	CV	77.39	77.86	78.75	86.17
	CD	22.61	22.14	21.25	13.83
DENA	CV	80.68	84.57	83.93	97.88
	CD	19.32	15.43	16.07	2.12
BrBA	CV	118.75	104.80	104.42	105.00
	CD	–	–	–	–

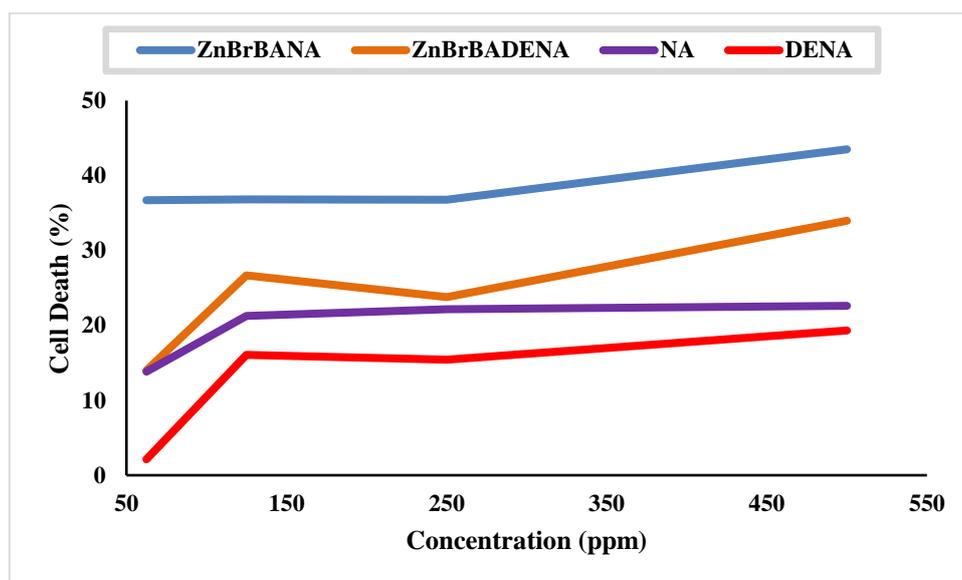


Figure 3. Change in the cell death according to the increasing concentrations

It was determined that DMSO used as solvent did not cause any cell death. Similarly, it was also found that 2-bromobenzoic acid, which is the primary ligand, did not have a cytotoxic effect on lymphocyte cells. NA, the co-ligand in crystal structure of ZnBrBANA, was cytotoxic at all concentrations. While the other co-ligand DENA did not cause cytotoxicity at 62.5 ppm, an average of 16.94 % cell death occurred in the 125-500 ppm concentration range. Cytotoxicity of ZnBrBANA and ZnBrBADENA complexes on lymphocyte cells was compared to each other and to the starting compounds (Figure 4-6). According to the results obtained, both complexes

caused cytotoxicity on these cells at the concentration range of 62.5-500 ppm. According to the results obtained, both complexes caused cytotoxicity on these cells at the concentration range of 62.5-500 ppm. ZnBrBANA complex is more toxic at 500 ppm than other concentrations. There is no statistically significant difference at concentrations of 250, 125 and 62.5 ppm.

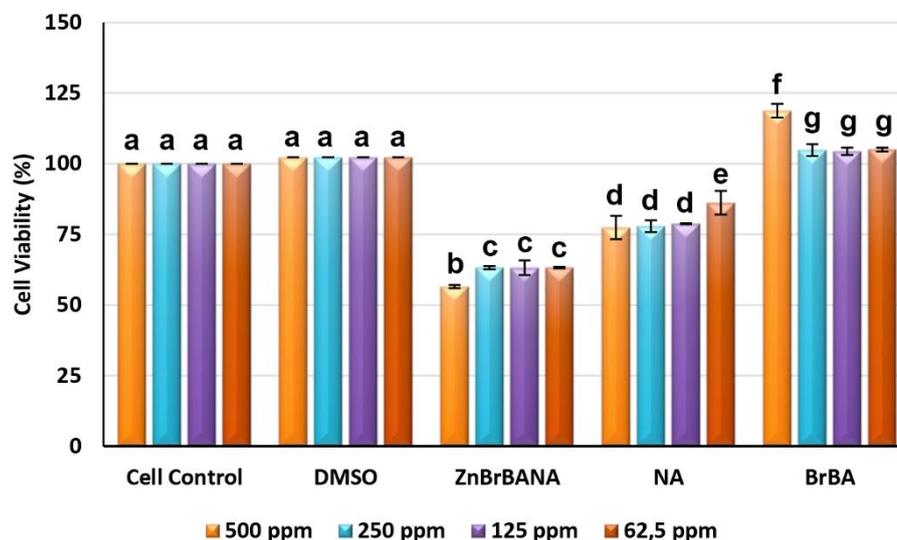


Figure 4. Comparison of cytotoxicity of ZnBrBANA complex with the starting compounds (Different letters on the columns were significantly different at $p < 0.05$)

Similarly, the ZnBrBADENA complex showed the highest toxicity at 500 ppm concentration. There was no statistically significant difference between the cell death ratios at 125 ppm and 250 ppm of this complex. ZnBrBADENA at 62.5 ppm was determined to cause less toxicity. The cytotoxicity of both complexes increases with the increasing concentration. The toxicity of the ZnBrBANA and ZnBrBADENA complexes compared to each other, the complex containing the DENA ligand was found to be less toxic. It is clear that this is related to the toxicity of the co-ligands contained in the complexes. In some studies, it has been reported that the toxicity of compounds that cause approximately 10 % cell death in normal cells can be neglected and these compounds can be used as anticarcinogenic agents (Bhattacharyya et al., 2019; Nashre-ul-Islam et al., 2019). Since the ZnBrBADENA complex causes 13 % cell death at 62.5 ppm, it is thought that it could be used as an anticarcinogenic agent in future studies. There are few studies in the literature on the determination of cytotoxicity of zinc arylcarboxylates with N-donor ligands on lymphocyte cells.

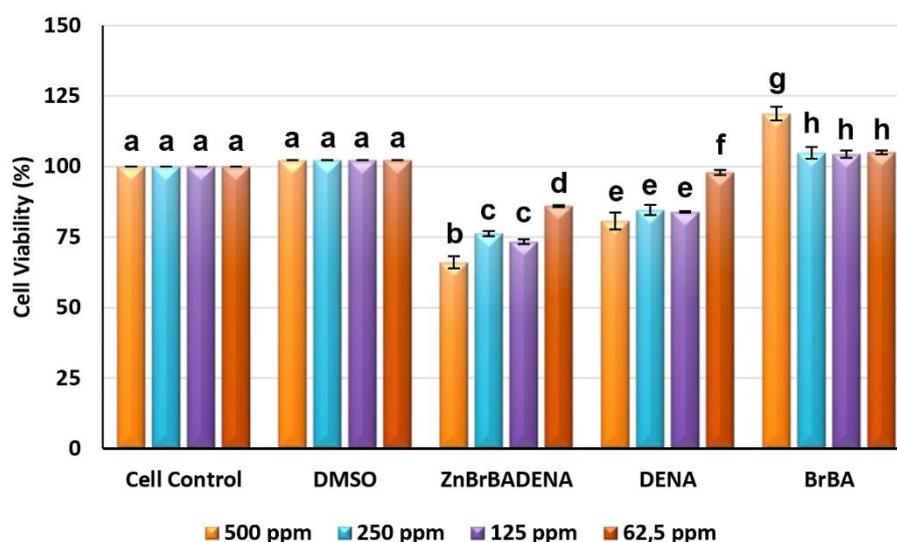


Figure 5. Comparison of cytotoxicity of ZnBrBADENA complex with the starting compounds (Different letters on the columns were significantly different at $p < 0.05$).

There is only one study in the literature on the determination of the cytotoxicity of zinc arylcarboxylates with pyridine derivative complexes on normal lymphocyte cells. It was reported that the zinc 2-fluorobenzoic acid nicotinamide complex decreased cell viability with increasing concentration at the 1.17-18.67 mM concentration range (Ozturk and Akbaba, 2019). In addition, it was determined that zinc 2-fluorobenzoate nicotinamide complex caused 13.80-93.58 % cell death in the concentration range of 1250-20000 ppm. Cell death caused by this complex at 10000 ppm is less than the toxicity caused by ZNBrBANA and ZnBrBADENA complexes at 500 ppm (Ozturk and Akbaba, 2019). ZnBrBANA and ZnBrBADENA complexes used in this study caused 43.5 % and 33.97 % cell death at 500 ppm, respectively. Generally, although metal complexes are recommended as an anticarcinogenic drug, their cytotoxic effects on normal cells are not investigated. The two complexes used in this study are not recommended to be used as an anticarcinogenic drug because they cause toxicity on lymphocyte cells at high concentrations (500 ppm).

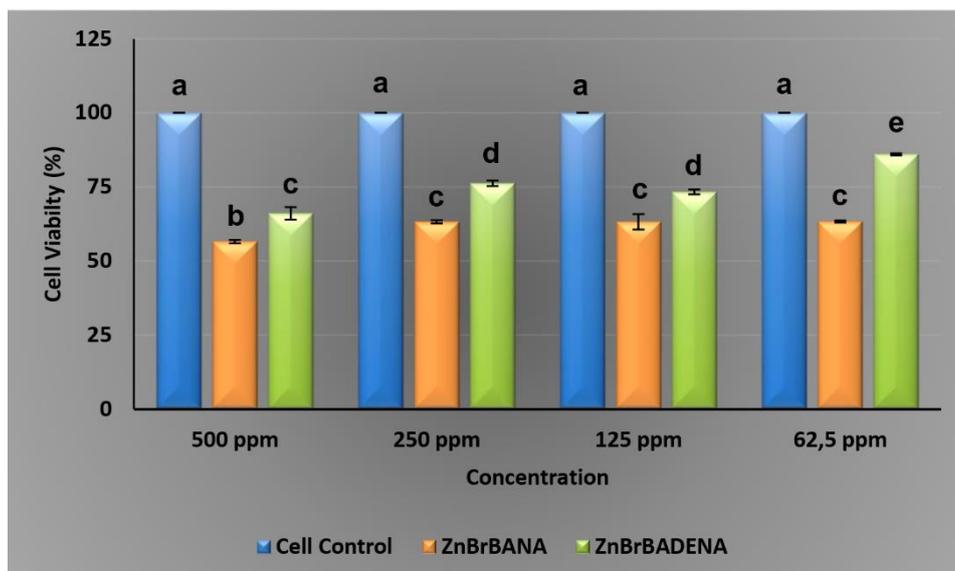


Figure 6. Comparison of cytotoxicity of ZnBrBADENA and ZnBrBANA complexes with cell control (Different letters on the columns were significantly different at $p < 0.05$).

4. CONCLUSION

It has been supported by many studies that metal arylcarboxylates are biologically active compounds in the last three decades. Although these compounds are recommended for use as new drug active ingredients, they must pass many tests for their active use. In this context, it is essential to know the cytogenotoxicity of the compounds. In many studies, a limited number of studies have been carried out on the toxicity of these compounds, which are stated to be effective against cancer cell lines, on normal cells. In this context, in this study, the cytotoxicity of Zinc (II) 2-bromobenzoate nicotinamide/N,N'-diethylnicotinamide complexes on lymphocyte cells in the concentration range of 62.5-500 ppm was investigated by MTT test. The ZnBrBANA complex was found to be toxic at all concentrations and the ZnBrBADENA complex caused moderate cell death at other concentrations, except 62.5 ppm. The cytotoxicity of the complexes increases with increasing concentration. At 62.5 ppm, the ZnBrBADENA complex is negligible toxic. We propose further studies to evaluate this compound, which causes 13.99 % cell death at this concentration, as DNA binders, tumor photosensitizers, antidiabetic, antifungal, antioxidant and antibacterial agent.

ACKNOWLEDGEMENTS

I would like to thank Füreya Elif Öztürkkan and Hacali Necefoğlu for their contribution in the supply of the compounds..

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